

HACKING HIV: CREATING AND ASSESSING A NOVEL CTL-BASED HIV-1 VACCINE

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Immunity Project
Waag Society, Amsterdam, NL
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IMMUNITY PROJECT

Immunity Project is a non-profit initiative dedicated to developing an Open-Access HIV vaccine.

We are a non-profit initiative of Flow Pharma, Inc. (Business hack)



EPIDEMIOLOGY

Global summary of the AIDS epidemic | 2013

Number of people living with HIV in 2013	Total	35.0 million [33.1 million – 37.2 million]
	Adults	31.8 million [30.1 million – 33.7 million]
	Women	16.0 million [15.2 million – 16.9 million]
	Children (<15 years)	3.2 million [2.9 million – 3.5 million]

People newly infected with HIV in 2013	Total	2.1 million [1.9 million – 2.4 million]
	Adults	1.9 million [1.7 million – 2.1 million]
	Children (<15 years)	240 000 [210 000 – 280 000]

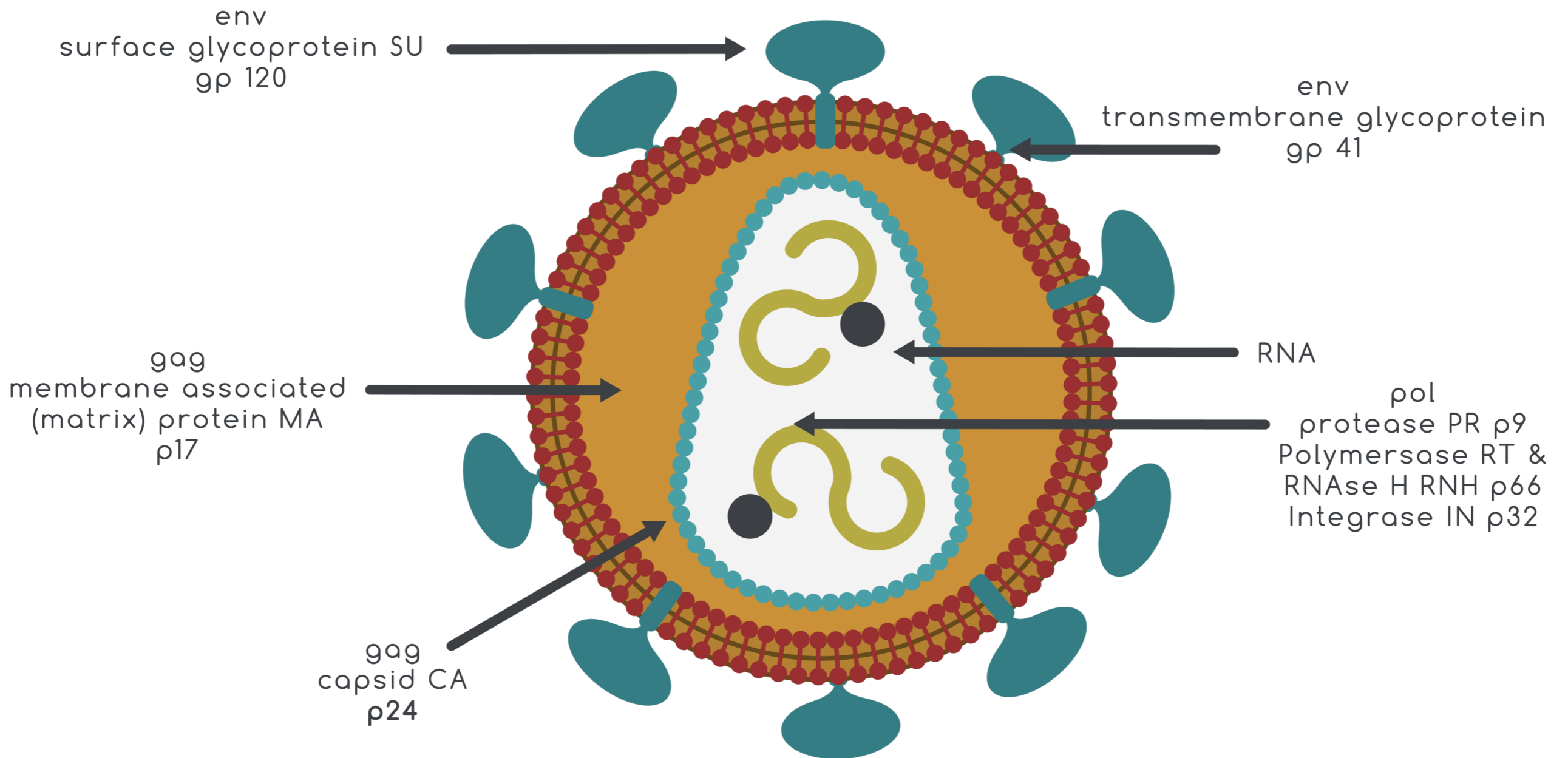
AIDS deaths in 2013	Total	1.5 million [1.4 million – 1.7 million]
	Adults	1.3 million [1.2 million – 1.5 million]
	Children (<15 years)	190 000 [170 000 – 220 000]

*Data from WHO, UNAIDS and UNICEF

More than 1.1 million people in the United States are living with HIV infection, and almost 1 in 6 (15.8%) are unaware of their infection.



THE VIRUS



IMMUNE RESPONSE TO HIV

- **Humoral** - antibody responses to virus are difficult due to high mutation frequency (old vaccines)
- **Cellular** - the infected body makes CD8+ CTL responses to viral infection, but these cells are down regulated during infection.
- Unless you're a **controller!**



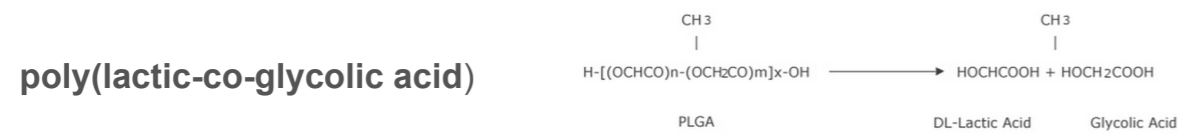
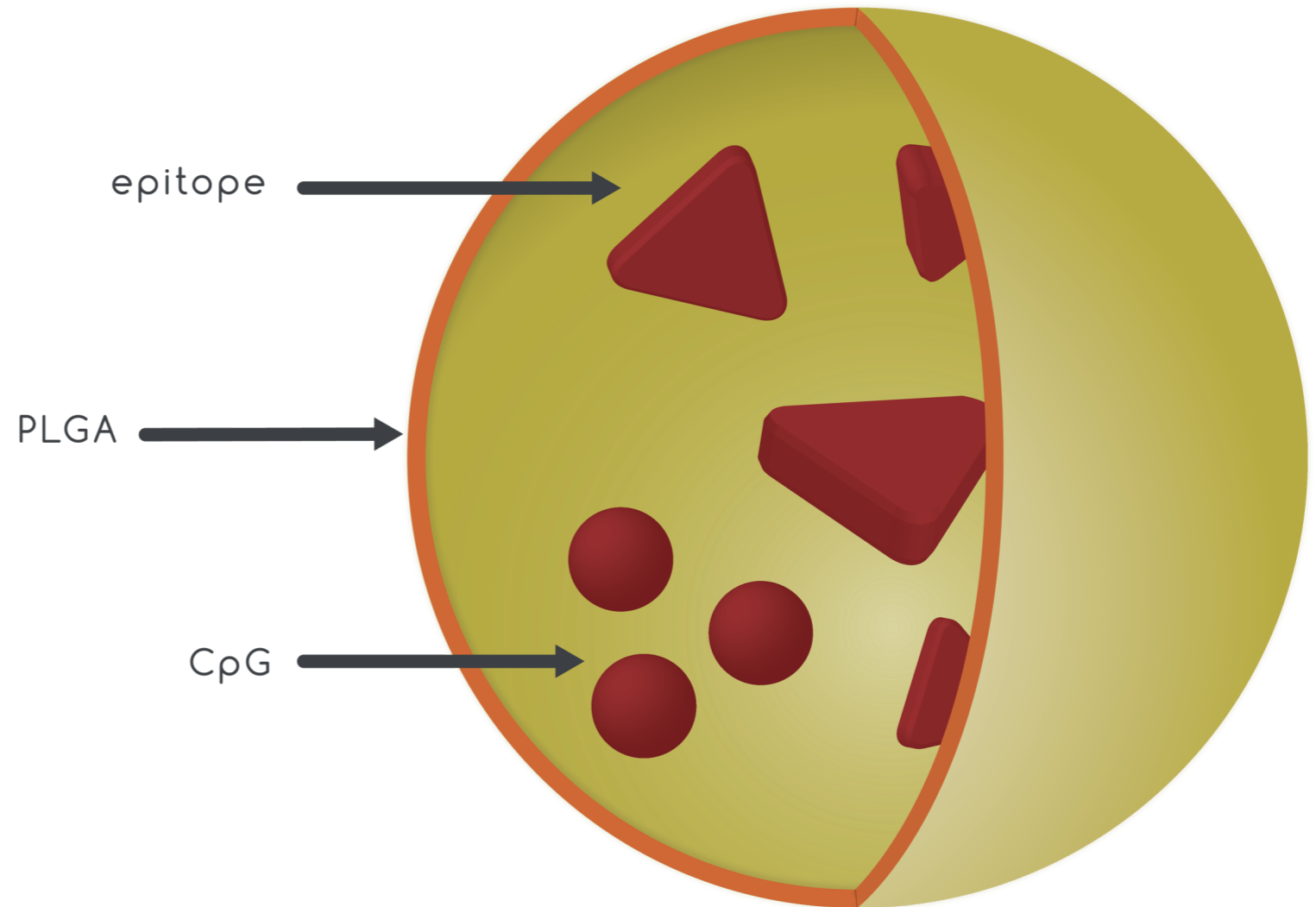
CONTROLLER

- **Controllers** have the natural ability to prevent advancement of HIV into AIDS.
- This is accomplished by the production of T cells with the ability to kill HIV infected cells (Killer T Cells).
- All people with HIV produce killer T cells, but in **non-controllers**, these cells are deleted by HIV-programmed cells.
- Different from **slow progressors**.
- We want to turn everyone into controllers!



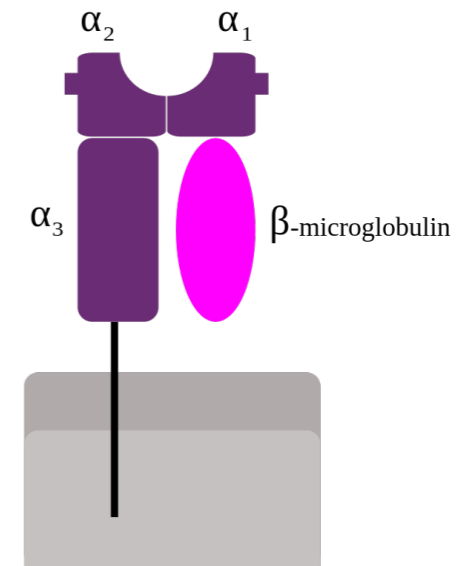
THE VACCINE

Component	Quantity
PLGA	98.5%
CpG	0.25%
Peptide	0.05% (100ng/mL)
Mannose	1.25%

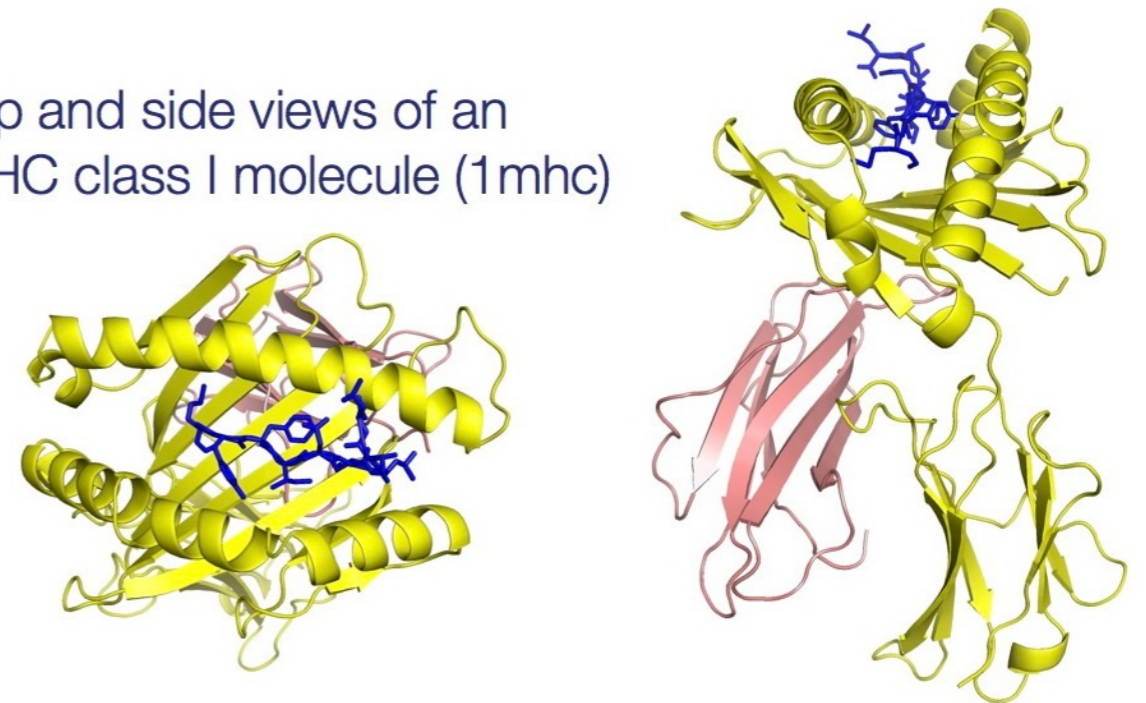


HACKING CTL PEPTIDES

- Identification of immunogenic proteins
- Identification of peptides that possibly bind to Class I molecules (based on HLA restriction)
- Creating databases of peptide binding (syfpeithi.de)
- Designing peptides for inclusion in our vaccine



Top and side views of an MHC class I molecule (1mhc)



A2, B57 AND B44

- Based on these databases, we chose two peptides specific for class I HLA: SL09 (SLYNTVATL) and KF11 (KAFSPEVIPMF)
- We performed a SYFPEITHI analysis to determine the binding probability of HIV-1 derived epitopes to class I molecules used in our studies.

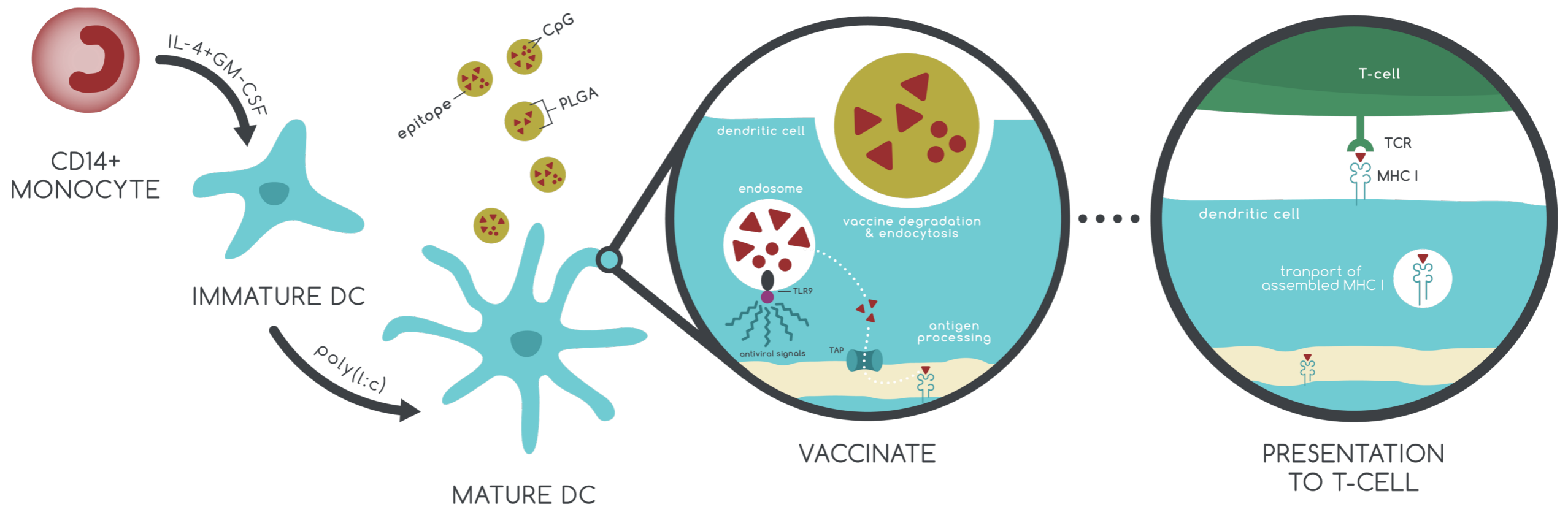
Haplotype	SL09 score	KF11 score	Positive Control Score
A2	31	9	31
B44	14	20	17 (228-239, vimentin)



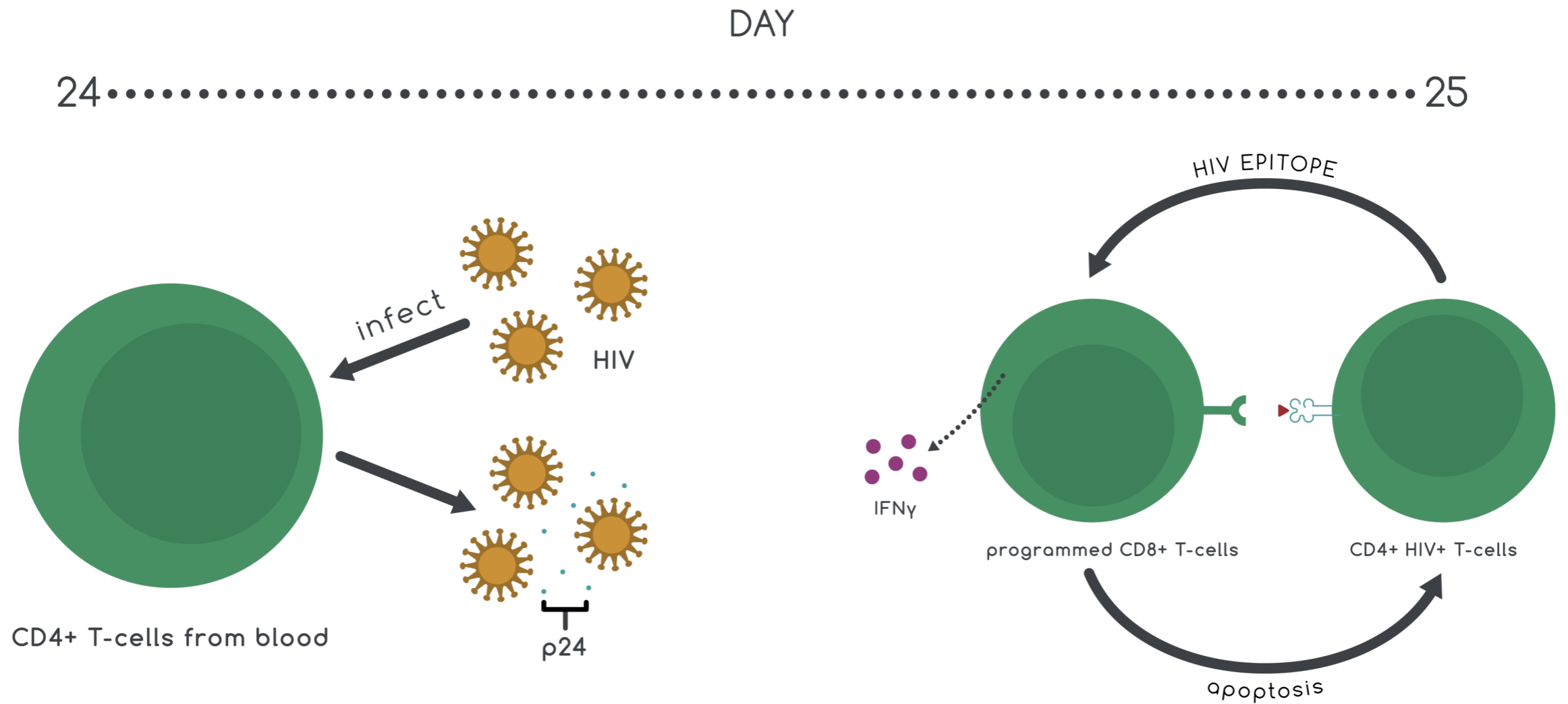
IN VITRO ASSAY (PART I)

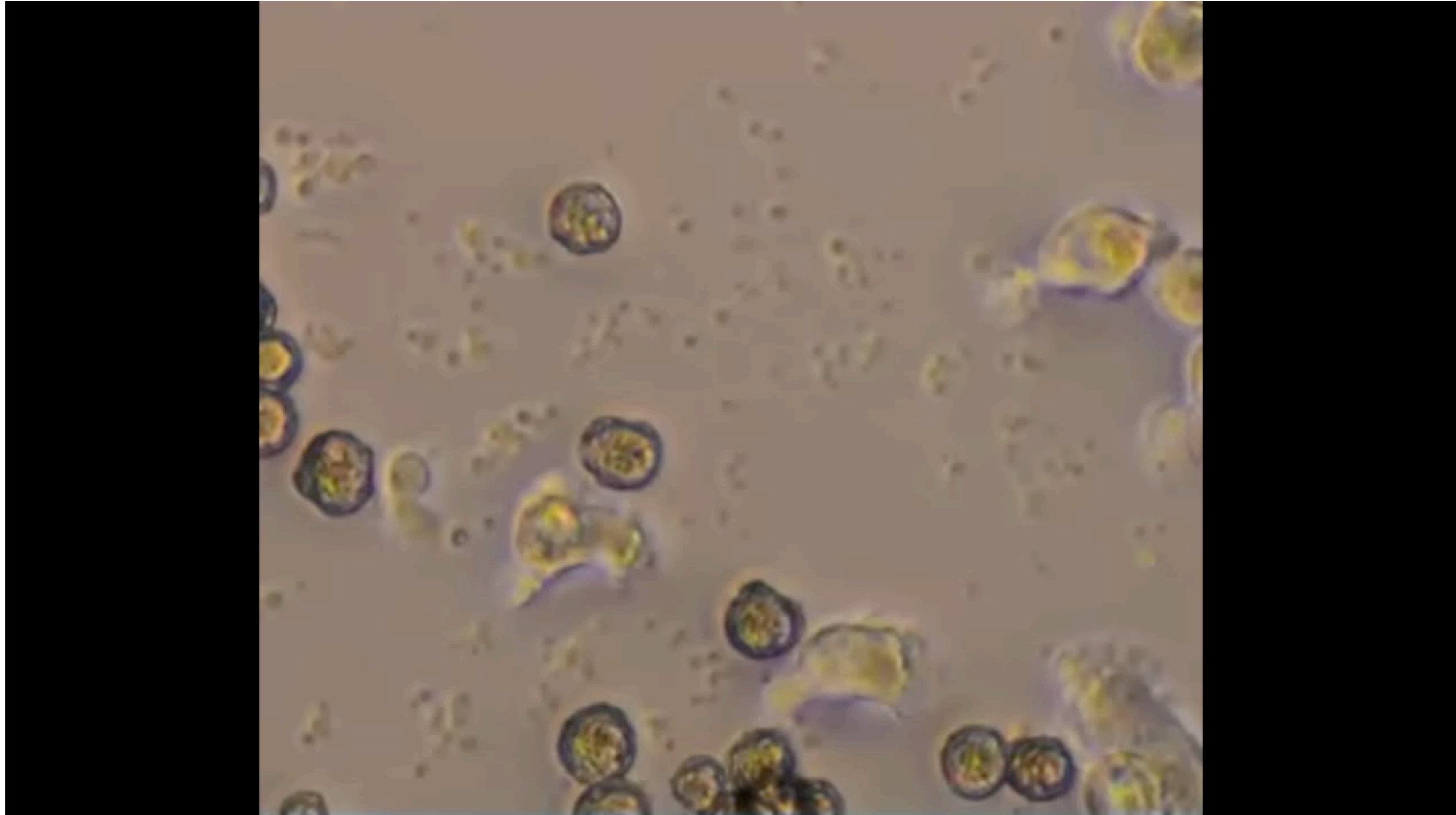
DAY

0 5 6 7 8



IN VITRO ASSAY (PART II)

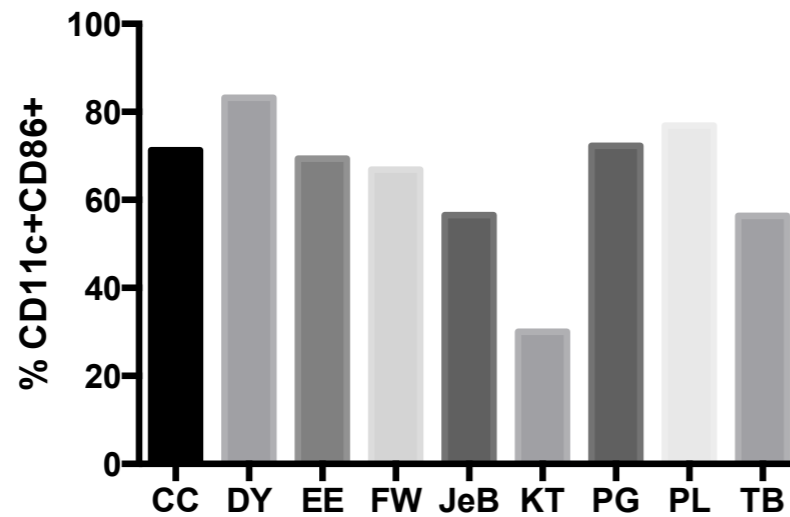




DENDRITIC CELL UPTAKE

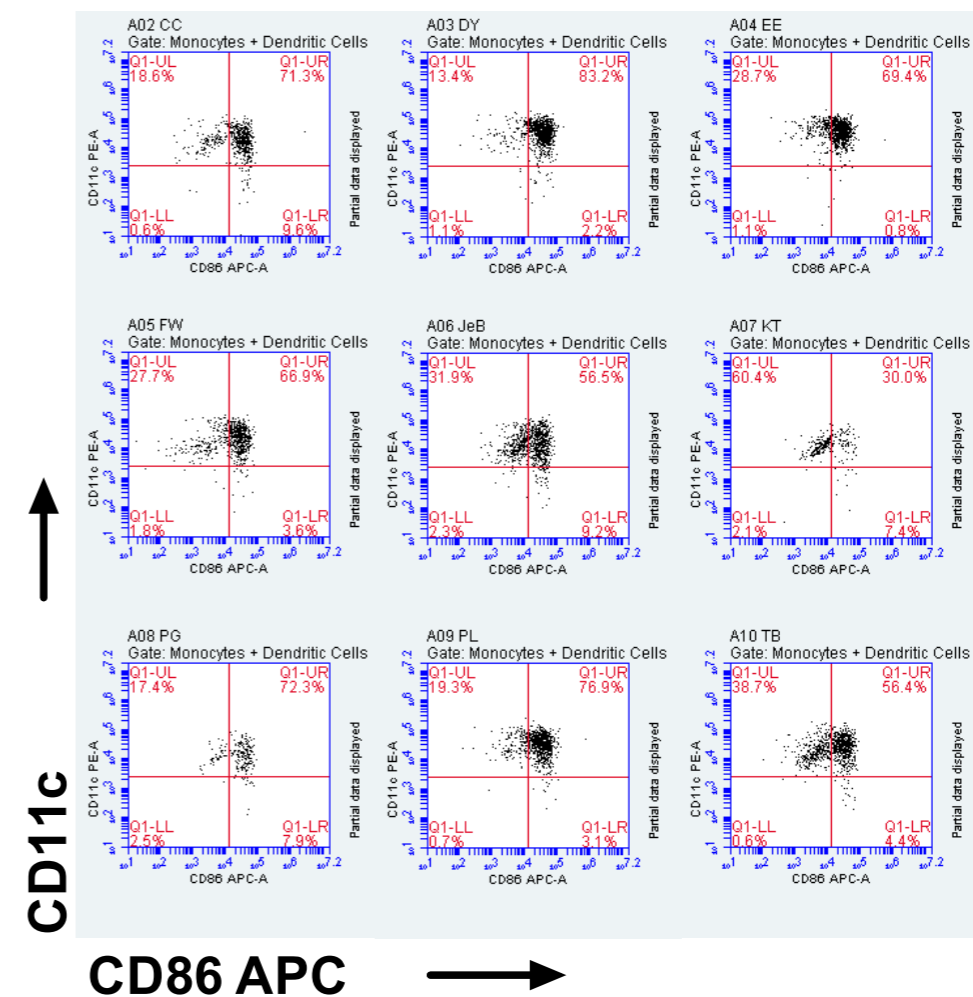


DENDRITIC CELL MATURATION



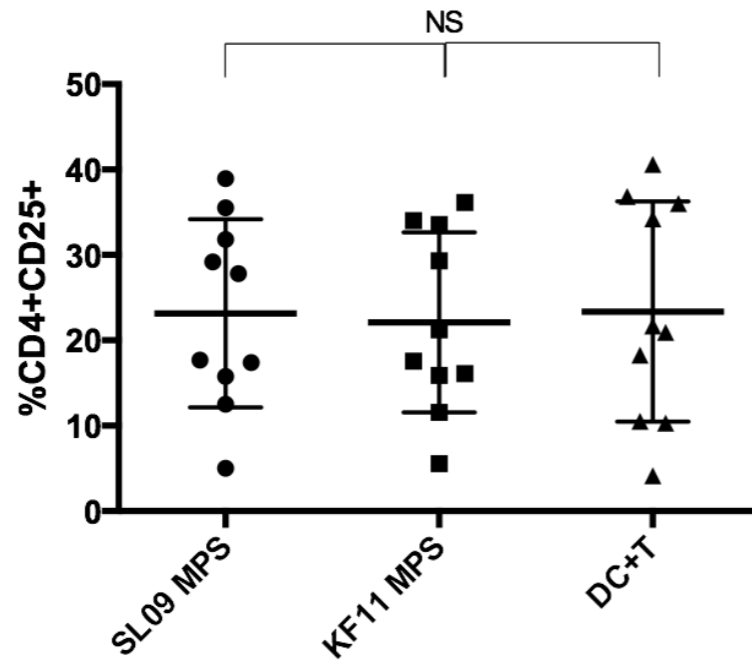
	CC	DY	EE	FW	JeB	KT	PG	PL	TB
CD11c+CD86+	71.3	83.2	69.4	66.9	56.5	30.0	72.3	76.9	56.4

- Flow Cytometry

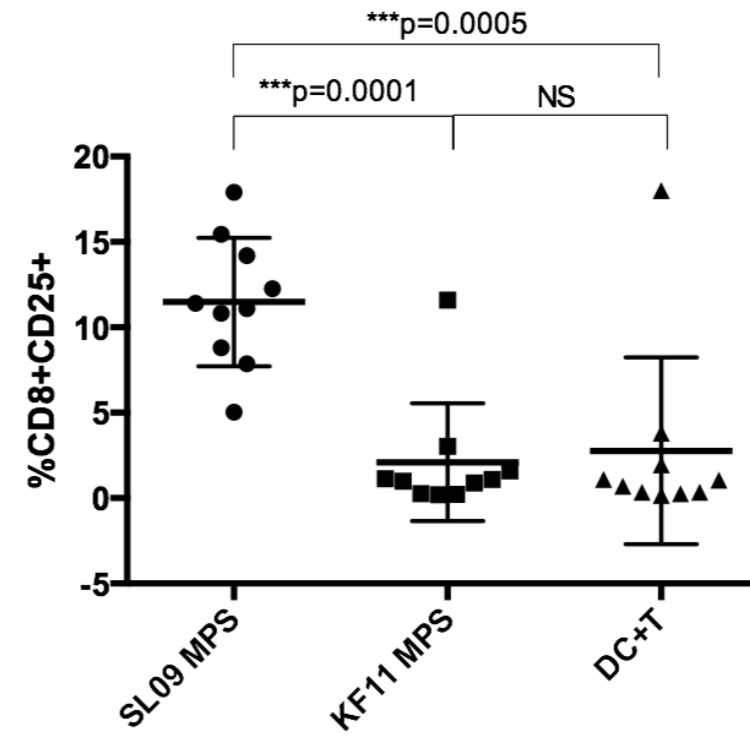


ACTIVATION

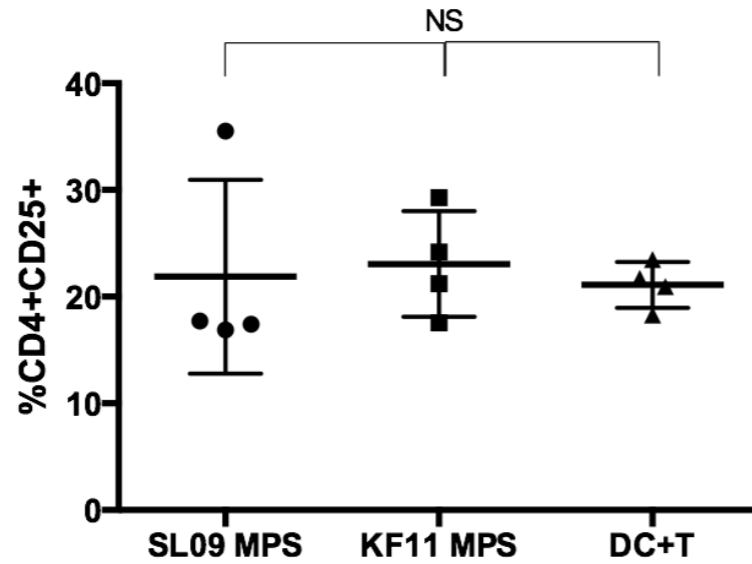
a)



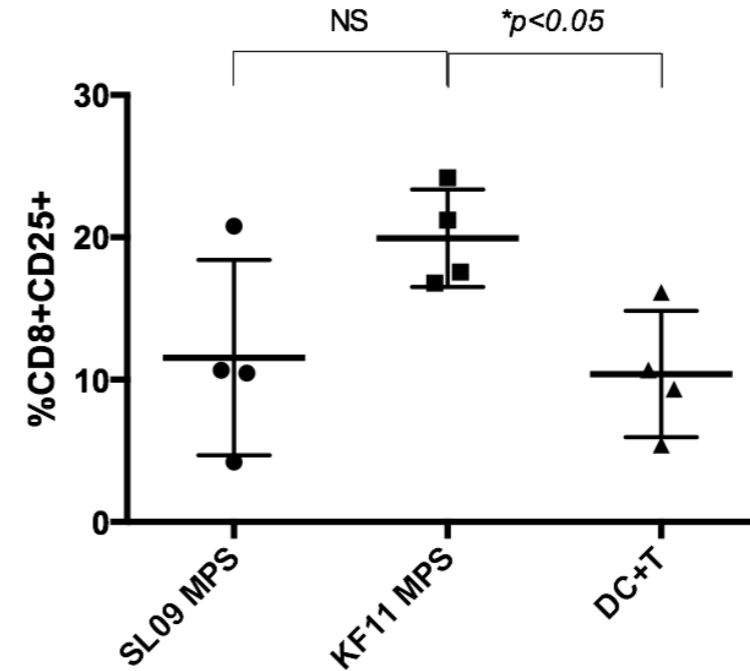
b)



c)

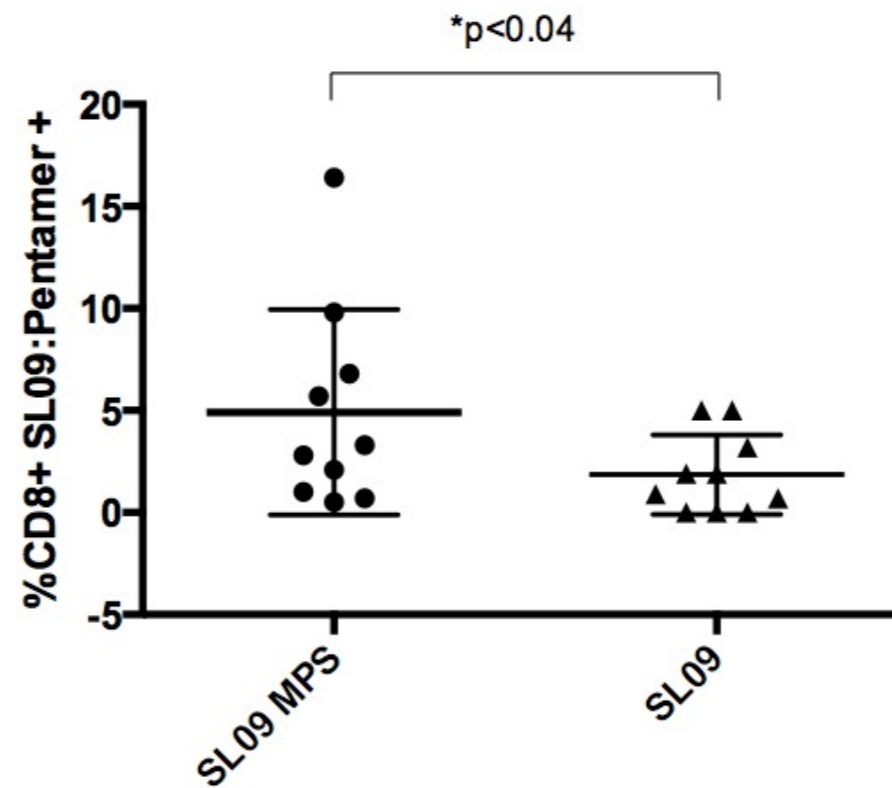


d)

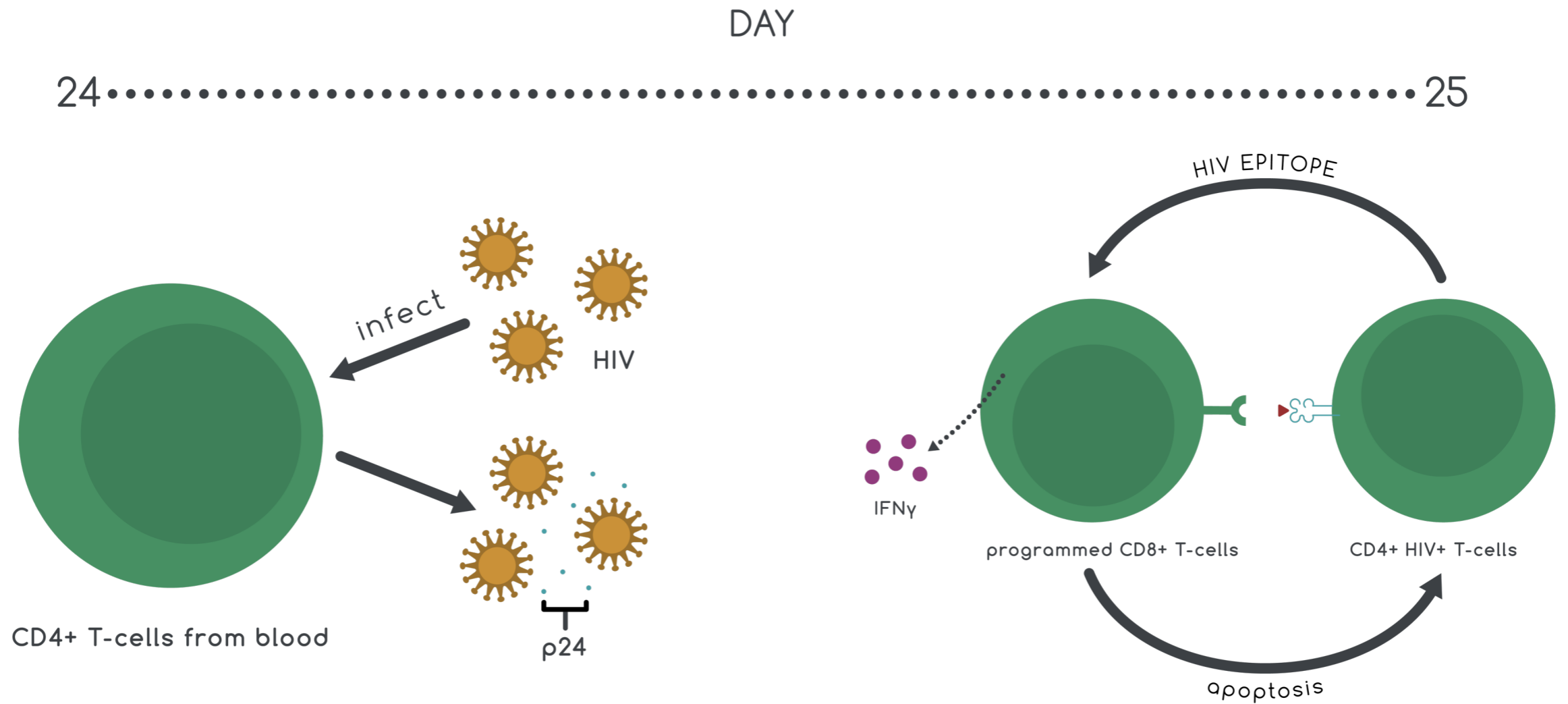


TCR UPREGULATION

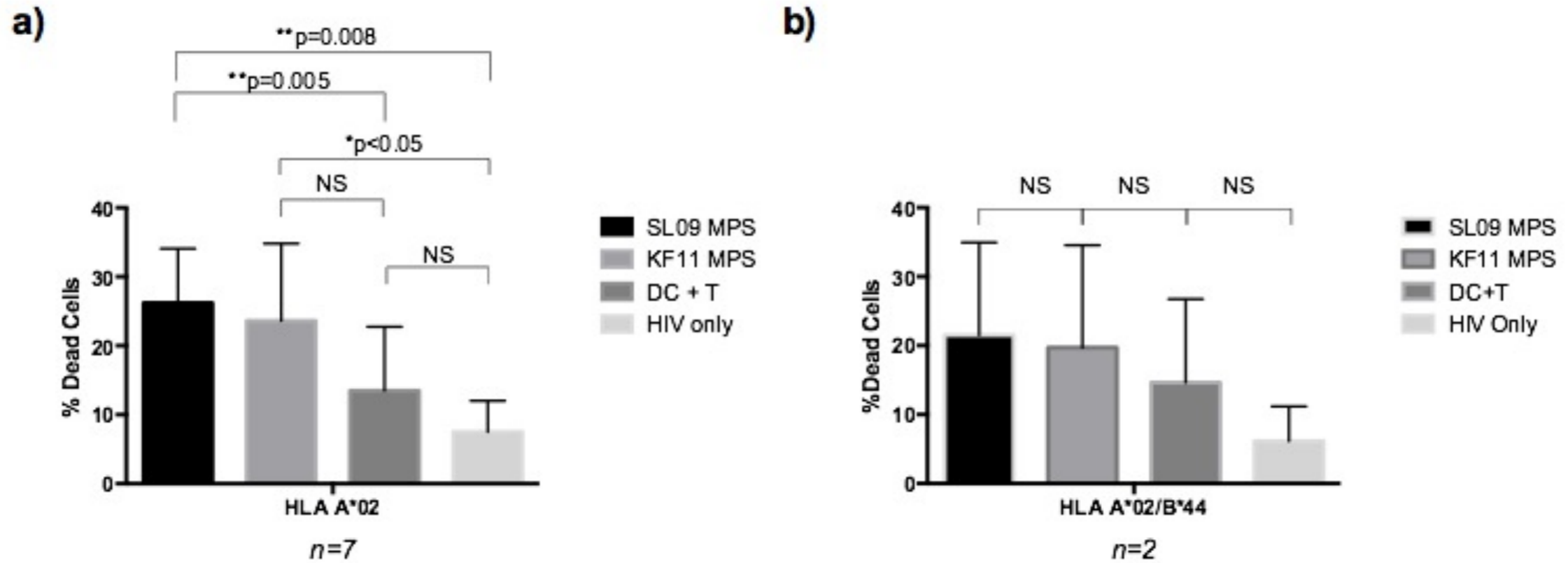
- On day 15, programmed CD8+ T cells were assessed for their expression of TCR that recognize SL09 in the context of HLA A*02 using A2:SL09 Pentamer (ProImmune).



IN VITRO ASSAY (PART II)



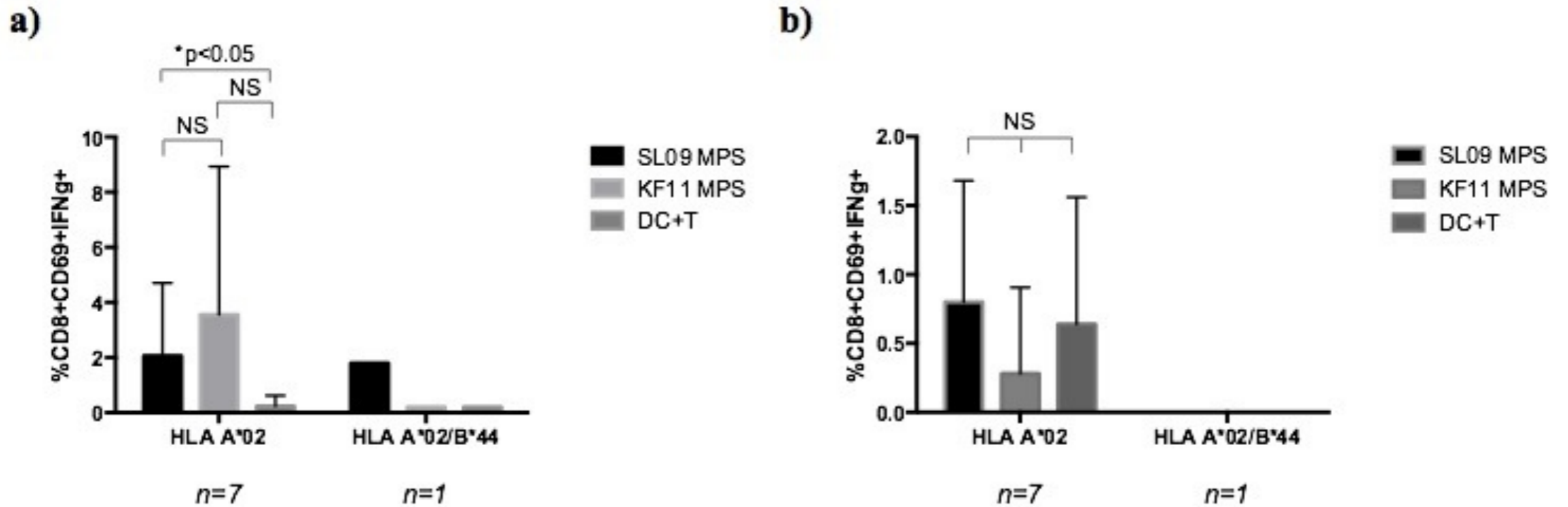
CTL-MEDIATED KILLING



- HIV-infected autologous CD4+ T cells were co-cultured with CD8+ T cells 5:1 (E:T)
- CD4+ T cells were assessed for apoptosis and death.
- CD8+ T cells were assessed for their ability to produce IFN γ in the presence of targets.



INTERFERON GAMMA

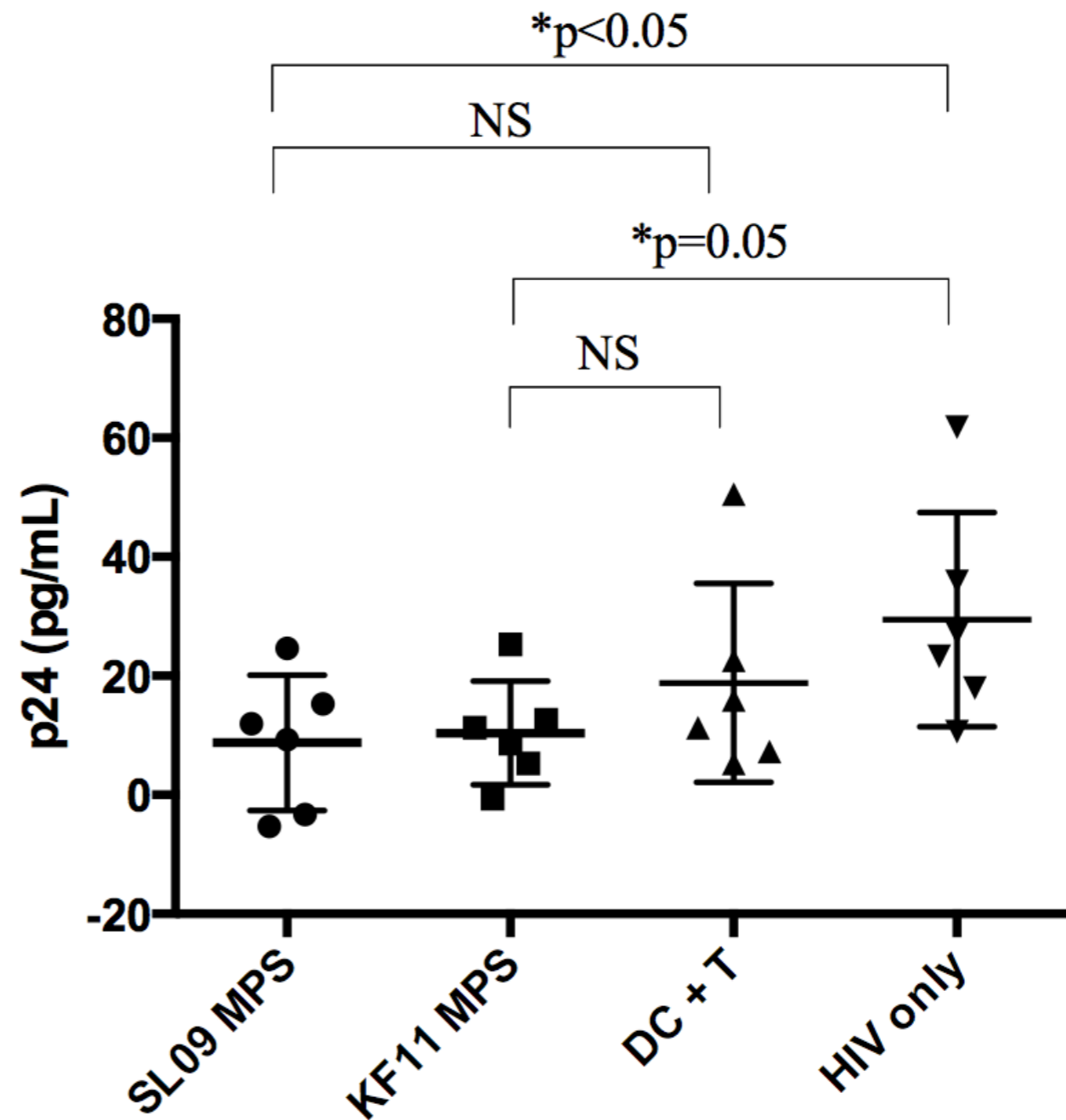


- CD8+ Effector T cells were co-cultured with autologous HIV+ CD4+ T cell targets and assessed for their production of Interferon gamma (IFN γ) on day 30.



P24 SUPPRESSION

- p24 ELISA was performed to assess the amount of HIV in the supernatant of CTL assay co-cultures.
- A decrease in p24 in tissue culture supernatant means that the T cells are effective at inhibiting HIV.



FUTURE DIRECTIONS

- We are designing pre-clinical experiments in Monkeys.
- We are working on clinical trials...
- We need a High Throughput Assay (that doesn't take 30 days to perform).



DANK JE WEL!

- Dank je wel Pieter van Boheemen en de Waag Society!
- Co-Founders:
 - Dr. Reid Rubsamen
 - Naveen Jain
- Scientists
 - Charlie Herst
 - Vikram Paranjpe (former Intern)
- Interns:
 - Rohun Patel
 - Hannah Hoban
- **COMMUNITY!**

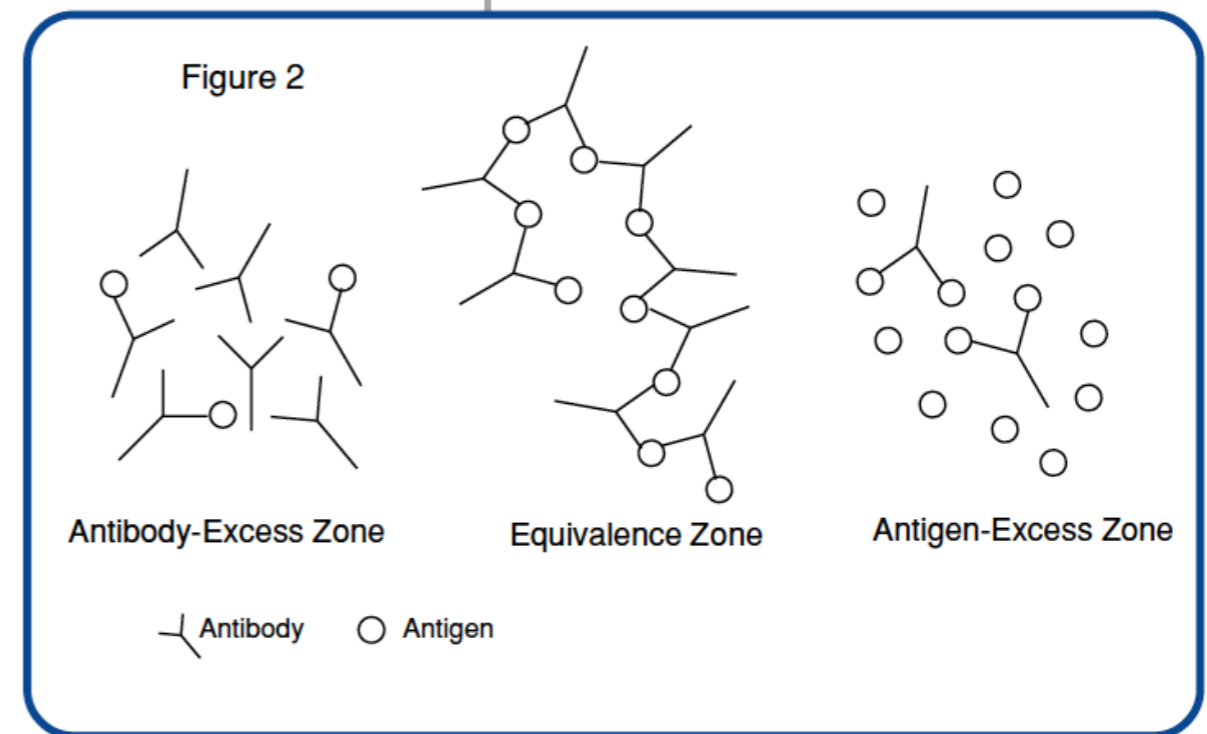
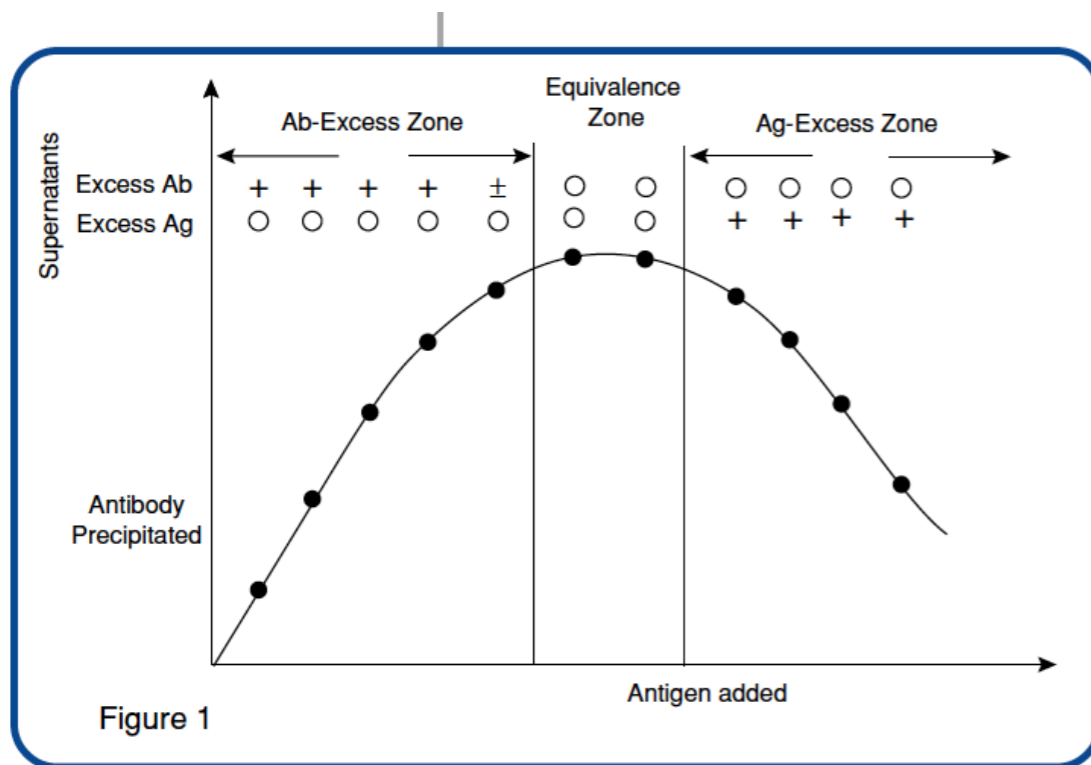


TODAYS EXPERIMENT

- Ouchterlony (Double Diffusion)
 - A test for reactivity of an **antigen** (protein) and **antibody**.
 - Precipitation reaction occurs and determines whether or not the antibody recognizes antigen.
 - This assay can be used in the field to determine whether or not a patient has come into contact with a virus containing a **known** antigen.



PRECIPITATION



PRECIPITATION IN TWO DIMENSIONS

- Add antigen and antibody to wells and assess precipitation....

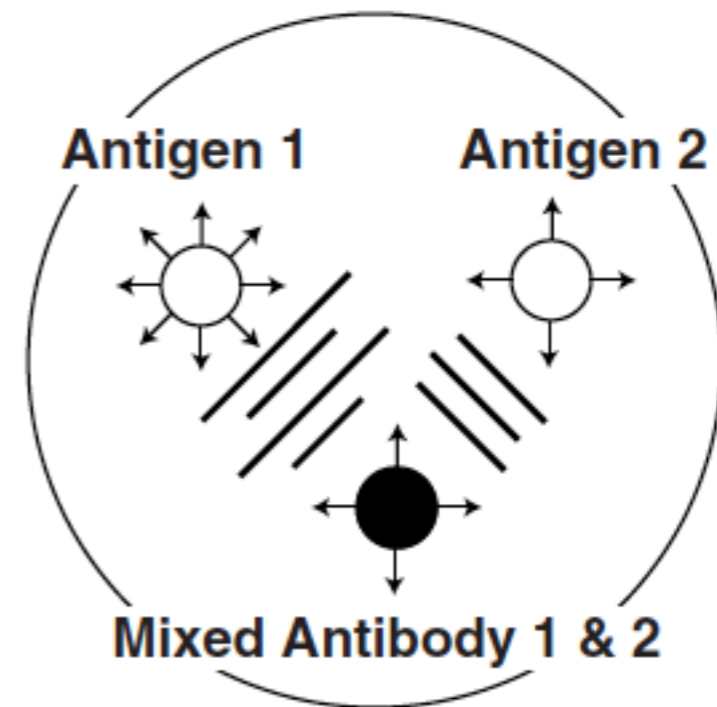


Figure 3



TODAY'S EXPERIMENT (CONT.)

- The Ouchterlony procedure allows us to measure different **reactivities**.

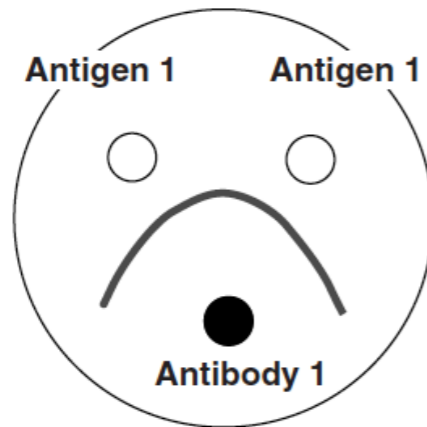


Figure 4:
Reaction of Identity

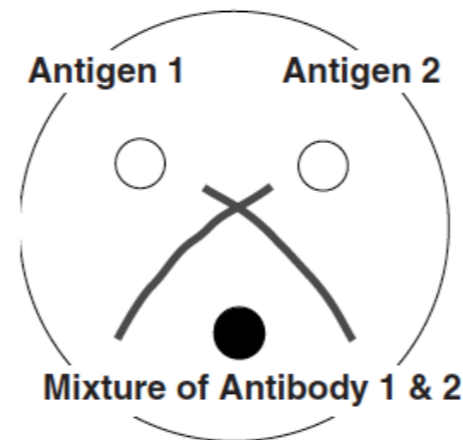


Figure 5
Reaction of Non-identity

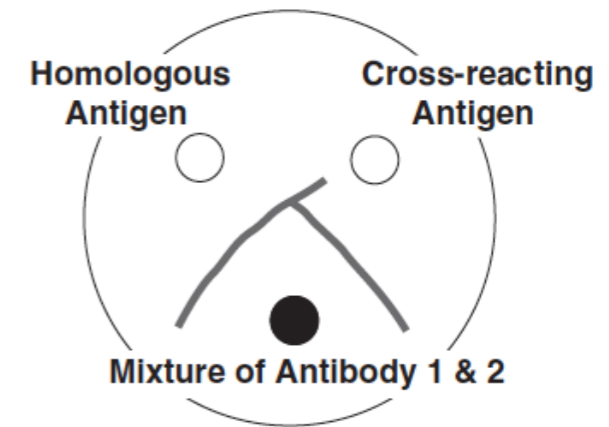


Figure 6
Reaction of Partial
Identity



PROCEDURE

- Pour plates (5mL of molten 1% agarose in Borate Buffer, pH 7.4)
- Allow to cool (5-10min)
- Punch holes in each plate (x3) and remove plug.
- Add antigen and antibody to each plate.
- Incubate overnight at 37C, in humid chamber.



PLATE SETUP

Tiny Holes

10uL of antibody
10ul of antigen

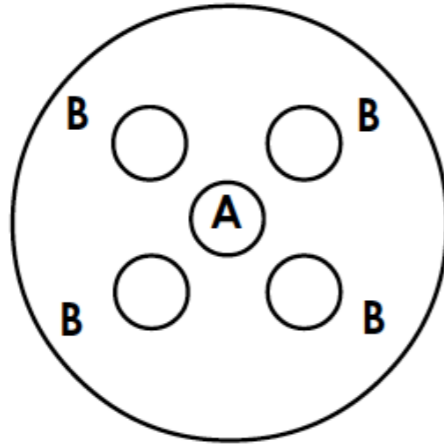


Plate 1 Center well: antiserum to the fluid containing antibodies (Tube A)
Left upper well: Whole serum (Tube B)
Right upper well: Whole serum (Tube B)
Left lower well: Whole serum (Tube B)
Right lower well: Whole serum (Tube B)

Large Holes

30uL antibody
30uL antigen

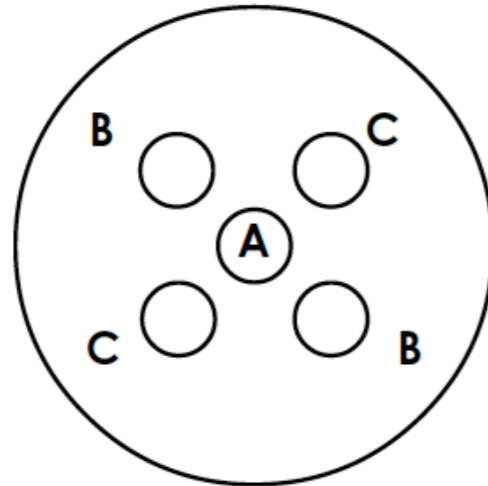


Plate 2 Center well: antiserum to the fluid containing antibodies (Tube A)
Left upper well: Whole serum (Tube B)
Right upper well: albumin (Tube C)
Left lower well: albumin (Tube C)
Right lower well: Whole serum (Tube B)

Plate 3 Center well: antiserum to the fluid containing antibodies (Tube A)
Left upper well: IgG (Tube D)
Right upper well: albumin (Tube C)
Left lower well: albumin (Tube C)
Right lower well: IgG (Tube D)

